

Imaging Sciences Centre

Novel MRI Active Gadolinium Liposomes for Cell Labelling

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Introduction

 Liposomes are artificially constructed spheres of amphipathic lipids that self assemble to form bilayers which enclose an aqueous compartment in the presence of water.

 The properties of liposomes can be altered based on adjusting the physical parameters of the constituent lipids (i.e. charge, head group, tail group). The aqueous interior can be loaded with a variety of molecules (nucleic acids or drugs) to render therapeutic functionality to the liposomes.



 Liposomes offer potential as diagnostic imaging agents given their biodistribution versatility; the incorporation of Gd lipids into a liposome formulation produces paramagnetic liposomes that can be used as targetable contrast agents in MRI.



MRI Analysis

•	Figure 5. Phanlorm MI Gd. DOTA, DSA with cc Coronal T, weighted in following parameters: 15 ms, FOV = 70 × 70 = 10, and matrix = 25 sequence used to obt T, values with 11 TR's	A images of ontrols. mages with the TR = 700, $TE =cm^2, averages\delta x 256 (semsain\phi) 4.7T.$
Sample no.	Content	T ₁ (ms)
1	0.5 mM Gd.DOTA.DSA in H ₂ O	400.1 ± 15.92
2	0.5 mM DOTA DSA in H ₂ O	2356 ± 112.588
3	Magnevist [®] in H ₂ O (equivalent Gd atom content as sample 1)	371.0 ± 4.321
4	H ₂ O	2750 ± 121.8

Figure 6 weighted	2 3 4 5 MR image of cell experiment. S images with the following param	$\begin{array}{c} 6 \\ \text{agittal } \mathbf{T}_1 \\ \text{neters: TR} = \end{array}$			
700, TE = 15 ms, FOV = 60 x 60 cm ² , averages = 4, and matrix = 256 x 128 (sems sequence used to obtain T ₁ values with 11 TR's) 4.7T.					
Sample no.	Content	T ₁ (ms)			
6	Gd Liposome	311.4 ± 40.10			
5	Gd Liposome + HeLa cells	688.6 ± 53.60			
4	Control Liposome	2248 ± 212.0			
3	Control Liposome + HeLa cells	2018 ± 132.8			
2	HeLa cells	1998 ± 163.7			

Table 2. Phantom MRI studies of new Gd lipid.

Table 3. MR images of cell pellet and controls.

PBS

1958 ± 146.2

 The T₁ value of the Gd.DOTA DSA phantom compares excellently with that of the commercial contrast agent Magnevist[®], the uptake and relaxation efficiency of the Gd liposome is verified by the bright cell pellet image in tube 5 (figure 6).

Conclusion

 Efficient uptake of Gd liposomes into HeLa cells was shown. Phantom MRI analysis of Gd.DOTA DSA revealed this lipid to be as effective as the commercial contrast agent Magnevist[®].

MRI analysis of HeLa cell pellets labelled with Gd liposome produced a brightly visible mass
of cells; reducing their T₁ value relative to control cells by 66%, and toxicity was minimal.

Intracellular uptake of liposomes was verified by fluorescence and confocal imaging.

DNA transfection with liposomes formulated with Gd.DOTA DSA gave comparable

transfection results to the commercial transfection agent Trojene® (Avanti Polar Lipids inc. USA).

Aims and Objectives

• The aim of this project has been to design and synthesise novel Gd lipids and to formulate Gd liposomes for *in vitro* cellular MRI labelling, and *in vivo* tracking of transplanted cells.

 The uptake efficiency of these liposomes can be assessed by finding an optimised cationic liposome formulation and hence the correct balance between the ratio of cationic, neutral and Gd lipids. This is done by the incorporation of a fluorescence probe in the liposome formulation and subsequent fluorescence uptake assays. The cell viability and cytotoxicity is also investigated along with the transfection ability of the optimised formulation with plasmid DNA.

 Gd liposome pre-labelling and MRI visibility of functional cells such as neutrophils (lung inflammation models) and stem cells (brain stroke legions/cardiac infarcts) will be investigated. The transfection ability of Gd liposomes for gene correction will also be studied in relevant models.

In Vitro Investigations

 A series of cationic liposomes were formulated using various ratios of CDAN, DOPE, Gd.DOTA DSA lipids and a small amount of a fluorescent lipid (Table 1). Liposomes were incubated with HeLa cells and the amount of liposome and thus Gd lipid uptake into cells was determined from the fluorescence emission intensity of the cell lysate.

 In addition, cell viability (MTT assay) and cytotoxicity (LDH assay) was assessed in order to determine which liposome had the best uptake/toxicity profile.

• Using this criteria liposome 2. was selected for imaging studies.

Liposome no:	CDAN mol %	DOPE mol %	Gd.DOTA.DSA mo	1%
1	30	30		40
control 1	30	30	DOTA.DSA	40
2	40	30		30
control 2	40	30	DOTA.DSA	30
3	50	0		50
control 3	50	0	DOTA.DSA	50
4	20	20		60
control 4	20	20	DOTA.DSA	60
5	40	30	Gd.DOTA.Chol	30
control 5	40	30	Cholesterol	30



Figure 3. Plasmid DNA transfection with an optimal formulation.

id DNA transfe



DAN: cationic lipid; facilitates endocytosis





Figure 4. Lipids used in liposome formulations along with Gd Lipid.

Fluorescence Microscopy



Figure 7. Fluorescence Microscopy Results (40 x magnification) Left Image: phase contrast. Right image: Fluorescence

• The fluorescence microscopy results show that Gd liposomes are taken up into HeLa cells and the intensity of the fluorescence is quite high.

 The image on the right of liposome formulation no.2 shows that the liposomes are ubiquitously present inside the cytoplasm of the cell, and are aggregated around the outer nuclear membrane. This also confirms that the mechanism of cell entry is potentially via endocytosis.



Figure 8. Liposome uptake is perinuclear

Future Directions

 The cellular uptake of the Gd liposomes using primary cell lines and their cellular biodistribution *in vivo* following transplantation or transfusion will be investigated i.e. cell migration and trafficking.